

Enteric parasitic infections: From environmental enteric dysfunction to gut microbiota and childhood malnutrition

Review
Article

Hanan M Abou-Seri¹, Mohammad Abdalgaber², Fatima Zahran³

Medical Parasitology Departments, Faculty of Medicine, Ain-Shams University^{1,3},
Cairo, and Gastroenterology Department, Police Authority Hospital, Giza², Egypt

ABSTRACT

Malnutrition accounts for high morbidity and mortality in children. Specifically, linear growth failure (stunting) has long-term consequences. It affects approximately 25% of children under the age of five worldwide and has been linked to increased mortality, cognitive dysfunction, and productivity loss. Understanding that a significant proportion of stunting is not caused solely by lack of nutrition, diarrhea, and repeated exposure to intestinal infections implies that other factors must be analyzed to clarify continued growth faltering. Enteric parasitic infections are prevalent in many tropical environments and are linked to poor nutritional status and growth stunting. Environmental enteric dysfunction (EED) is an undefined syndrome characterized by inflammation, decreased absorptive capacity, and impaired barrier function in the small intestine. The dysfunction has been allied to malnutrition, as well as oral vaccine failure causing delayed development in children from poor resource settings. This syndrome might be triggered by unhygienic environmental conditions that cause frequent exposure to fecal pathogens and decreased nutrient absorption. Additionally, childhood malnutrition was strongly linked to changes in the gut microbiome. Motivation for the present review was the relatively obscure cumulative effect of repeated enteric parasitosis on nutritional status of children in developing countries from various perspectives. Accordingly, intestinal parasites infections, EED, and microbiota alteration are all likely issues that must be concurrently addressed. Addressing all these conditions would resolve the grave public health issues of malnutrition and infection susceptibility, and consequently, lead to application of behavioral and therapeutic policies among vulnerable pediatric populations.

Keywords: children; enteric parasites; environmental enteric dysfunction; gut microbiota; malnutrition.

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Corresponding Author: Hanan M. Abou-Seri, **Tel.:** +20 1006770522, **E-mail:** hanan.mahmoud27@hotmail.com

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INTRODUCTION

Child health is a prerequisite for the development of nations' futures. Therefore, maintaining optimal child health and growth is necessary for the economic development of countries^[1]. Child malnutrition is a major public health issue, particularly in many low and middle income countries. In approximately 45% of all deaths reported for children under the age of five, undernutrition is the underlying cause^[2]. Malnutrition is induced by a child's shortage of essential nutrients, and accounts for underweight, stunting, and even wasting^[3].

Many factors contribute to malnutrition. In particular, EED, is considered one of the leading causes of malnutrition. The pathway from EED to malnutrition and stunting is hypothesized to be through malabsorption and chronic inflammation, which arise from microbial and parasitic infections associated with an impaired gut barrier^[4,5]. Later, gut helminths, protozoa, and more recently, microbiota have all been linked to the multifaceted web of elements involved in undernutrition^[6].

Enteric parasitic infections are a significant public health issue, particularly in developing

countries. Worldwide, more than three billion people (mostly children) are believed to be infected^[7]. Soil-transmitted helminths (STH); namely, hookworms, *A. lumbricoides*, *T. trichiura*, and *S. stercoralis* are responsible for the majority of these infections^[8]. These STHs are ranked as the most crucial human parasitic infections in terms of disability adjusted life years (DALYs) lost; outranking other infections such as African trypanosomiasis, Chagas disease, and leprosy^[9]. *T. trichiura*, alone, infects an estimated 477 million individuals, with children recording the highest infection prevalence and intensity^[8]. Additionally, children under the age of five were found to be frequently infected with *Cryptosporidium* spp., *E. histolytica*, and *G. duodenalis* in Sub-Saharan Africa^[10].

Substantial evidence links between parasitic infection and poor nutritional status, in geographical areas where both are prevalent^[11,12]. Malnutrition renders the populace more vulnerable to parasitic diseases and *vice-versa*. Some nutrient deficiencies can impair the host's immune function, potentially increasing susceptibility to infections^[13]. Enteric parasitic infections, in particular, can impair the nutritional status by causing intestinal bleeding

and nutrient competition, resulting in nutrient malabsorption^[14]. These parasites may also lessen the benefits of food intake by causing anorexia, reducing fat absorption and protein utilization; as well as increased nutrient waste through vomiting and diarrhoea. Protein energy malnutrition (PEM), anemia, and other nutrient deficiencies are the result of these conditions^[14]. Moreover, chronic infections with intestinal parasites can interact with microbiota, also leading to malnutrition^[6].

Hence, the aim of the present review was to highlight a set of interacting pathways that might aid in elucidating the link between enteric parasitosis and malnutrition (stunting in particular) due to micronutrient deficiency, EED, and parasite-microbiota interplay.

Stunting

This impediment of development is a landmark of malnutrition resulting in reduced productivity in adulthood. The role of intestinal parasitic infections in stunting and even wasting were significantly observed among children with helminthic infections^[15]. In fact, *A. lumbricoides* is one of the most prevalent enteric geohelminths globally^[16], strictly linked to stunting among children^[17], with subsequent lagging of their linear growth and deterioration of their cognitive functions^[18]. Simultaneously, the etiology of malnutrition has been related to diarrhea-causing intestinal protozoan infections, where repeated diarrhoeal attacks were considered as a risk factor for stunting^[19,20]. In Bangladesh, stunted infants were at a higher risk of developing severe diarrhoea, particularly that linked to infection with *E. histolytica* and *Cryptosporidium* spp.^[21]. Children with asymptomatic and symptomatic cryptosporidiosis were found to gain less weight in the first month of infection, and once infected they are not liable to "catch up growth" and are predisposed to growth stunting^[12]. Nevertheless, the mechanisms underlying growth impairments after infection are complex, and most likely linked to EED^[21].

Micronutrient deficiencies

Vitamins A, C, D, E, B6, B12, folate, zinc (Zn), selenium, copper (Cu), and iron all have immunomodulatory properties that affect the course of an infection^[22]. Iron, Cu, Zn, and magnesium are essential for growth and development. They are implicated in multiple metabolic functions and are responsible for the integrity of the natural human defense mechanism. Their deficiency causes impaired immunity that exacerbates the nutritional status^[23]. Iron, in particular, is the most common deficient micronutrient and the leading cause of anemia worldwide, especially among infants^[24]. An intense correlation between iron deficiency anemia in children and parasitic infections was established owing to multiple risk factors, mainly fecal iron leakage caused by parasitic infections such as hookworms, schistosomiasis, and trichuriasis^[24]. Trichuriasis was

also linked to low intakes of macronutrients and micronutrients such as protein, energy, iron, thiamine, and riboflavin^[25].

Vitamin A deficiency and exposure to STH infections overlap vastly among children, especially in poor resource settings. They are implicated in major health and economic disabilities and have been addressed by public health measures for years^[26,27]. To overcome this deficiency, supplementation of deworming drugs with vitamin A might show potential synergistic health benefits^[28,29]. Analysis of various micronutrient supplements on intestinal parasitic reinfections showed that while Zn supplementation is ineffective, iron supplementation had a positive effect^[30]. In order to augment hemoglobin level and nutritional status in STH endemic areas, the World Health Organization (WHO) recommends regular-interval deworming of school children with anti-helminthic medications^[31]. Accordingly, academic performance improvement, child mortality decline, and raised economic productivity should occur^[31].

Environmental enteric dysfunction

1. Role of enteric parasitic infections in enteropathy:

A variety of environmental and host factors are thought to contribute to EED^[32]. The scarcity of specific disease knowledge regarding this pathological condition requires further research into its pathogenesis, complications, and possible treatments. This enteropathy shares common features with other gut chronic inflammatory conditions, including celiac disease and inflammatory bowel disease (IBD)^[33].

Repeated enteric infections in the first two years of a child's life can result in impaired intestinal absorptive function^[34]. It is thought that even an infection load that is inadequate to trigger diarrhea can cause EED^[35]. Human protozoal infections, such as cryptosporidiosis, microsporidiosis, cyclosporiasis, isosporiasis, and *Giardiasis*, in addition to STH can directly disrupt the intestinal barrier by binding to cell surface molecules, causing cell damage and apoptosis, or by disrupting tight junctions and cell cytoskeleton^[36]. Besides, the parasites elicit a strong innate mucosal immune response, including neutrophil activation and other cells that contribute to the intestinal lesions^[37]. The inflammatory response in the intestine is affected by the invasive ability of the parasite and host's immune status, which in turn will have an impact on parasite load, and duration of infection^[38].

Infection with *Cryptosporidium* spp. has been linked to increased gut inflammation and villus architecture loss; and murine models suggest that immune signalling in the gut may be interrupted, resulting in enteropathy and poor growth^[12]. *Blastocystis* spp. are opportunistic parasites with a negative impact on gut health in the presence of other pathogens. However, evidence of its pathogenicity in humans remains speculative and

inconclusive^[39]. *Blastocystis* spp. was found prevalent in patients with impaired intestinal barrier function and altered gut permeability. But the relation between *Blastocystis* spp. and EED remains inconclusive^[40]. It is unknown whether the presence of this parasite is an association due to decreased immunity or if it causes harm through EED. However, EED is believed to occur as a result of repeated exposure to fecal pathogens like *Blastocystis* spp.^[40].

The definitive diagnosis of EED is through gastrointestinal tract endoscopic and histopathological examination. Changes in the intestinal structure recorded in EED include blunt villi, increased crypt depth, reduced compartmental features, and increased shedding of epithelial cells^[41]. Moreover, increased cellular infiltrates with the predominance of lymphocytes and plasma cells also occur. Hence, the intestinal functions are affected by abnormalities of cell adhesion and tight junctions^[41]. Many non-invasive biomarkers were developed for EED, such as alpha-1 anti-trypsin, calprotectin, myeloperoxidase, and lactulose:mannitol (L:M) ratio. However, lack of validation hampered research, raising difficulties in assessing EED^[42]. It worth mentioning that L:M ratio was used as a biomarker for young children due to the practical and ethical concerns entailed by endoscopic biopsies. An association was also revealed between parasitic infection and higher L:M ratios, indicating increased intestinal permeability^[43]. Novel biomarkers are constantly sought, but finding a 'gold standard' test for assessment has not been achieved yet^[42].

Nutritionists are collaborating with experts in the fields of water, sanitation, and hygiene (WASH) in countries with high stunting rates, and their combined efforts are assisting the regional hygiene programmes^[32]. Essentially, improvement of WASH is a critical component of achieving global stunting elimination. Its development can disrupt the fecal-oral transmission pathway^[44]. However, there are few studies that specifically investigate baby WASH, which is designed to target maternal and newborn health during the first 1,000 days. They revealed its effectiveness in reducing stunting in children under 2 years^[44-46].

2. Role of extensive use of antibiotic in enteropathy:

Antibiotic treatment could counteract chronic exposure to environmental microbes and pathogens that cause enteropathy, thereby improving child growth in developing countries^[47,48]. However, antibiotic overuse may affect the gut microbiota, which plays an important role in pathogen regulation, resulting in a change in their diversity and increased susceptibility to harmful pathogens, and thus enteropathy^[47,48]. It was shown that patients who had previously received antibiotics were more susceptible to *Escherchia coli*, *Salmonella*, *Shigella*, and *Campylobacter* infections; as well as recurrent *Clostridioides difficile* infections^[48].

Overuse can also result in antibiotic resistance to many pathogens, including bacteria and parasites. Parasites may develop novel mechanisms to achieve drug resistance, which necessitates substantial efforts for combat^[48,49].

3. Role of food additives in enteropathy:

As indicated by Marion-Letellier *et al.*^[50], the increased consumption of ultra-processed food in developed countries, may be a significant cause of chronic diseases; although it is still unknown whether this association is due to the poor nutritional composition or the presence of food additives. The various additives reported to keep foods fresh or enhance their color, flavor, or texture include glucose, salt, emulsifiers, organic solvents, gluten, microbial transglutaminase. Additionally the authors indicated that also nanoparticles are increasingly used by the food industry to improve the quality of food, and warned that most of these additives can trigger gut inflammation causing tight junction dysfunction and increased intestinal permeability. For example, processed food was found to include high levels of sodium chloride, which can exacerbate colitis resulting in enteropathy or worsening of inflammatory bowel disease. Another element incorporated in processed food is aluminum, added from wrapping foils or cooking vessels. A high level of aluminum was found to worsen colitis. All of these factors indicate the role of food additives in causing gut inflammation, which can increase susceptibility to infection and enteropathy^[50]. Former studies had indicated that the opened tight junction could result in foreign immunogenic antigens entering and activating an autoimmune cascade causing autoimmune disease^[51,52]. Another statement implicated artificial emulsifiers and sweeteners in inducing dysbiosis associated with alteration of the intestinal barrier, activation of chronic inflammation, and abnormal immune response accelerating the onset of IBD^[53].

Parasite-microbiota interplay

Gut microbiota are a diverse and stable microbial community that is established at birth. This ecosystem performs a wide range of anti-infectious, anti-inflammatory, and immune-modulating functions, essential for intestinal homeostasis^[54]. Microbiota interact with the host on a molecular level by producing and responding to a variety of neurotransmitters and endocrine molecules that affect appetite, intestinal transit time, lipid and glucose metabolism^[55]. In a study conducted by Kane *et al.*^[56], malnutrition was linked to delayed normal development of the gut microbiota in early childhood, coupled with the disruption of the microbiota composition that can lead to a lack in vital roles for healthy growth, and/or an elevated risk of intestinal inflammation^[56].

Chronically malnourished children were observed to exhibit an immature microbiome, which was linked

to linear growth failure^[57]. Besides, since microbiota are required for the development of the mucosal immune system of the intestine, "dysbiosis," or altered microbiota composition, is thought to be involved in the development of EED and, as a result, malnutrition. This was conveyed by researchers who transplanted the feces of malnourished Malawian children to gnotobiotic mice, resulting in malnourishment among the murines^[58,59].

Studies documenting the interaction between parasites and gut microbiota showed that the direct or indirect modifications in the microbiota might be positive, neutral, or negative^[60,61]. Parasites can affect microbiota populations by stimulating an anti-inflammatory immune response caused by their attachment and secretion of molecules. These molecules improve barrier function by stimulating goblet cell hyperplasia and increasing mucus volume, thus reducing bacteria adherence to the epithelium^[6]. Several trematodes including *Schistosoma*, *Fasciola*, *Opisthorcis*, *Clonorchis*, and *Paragonimus* spp. secrete peptides known as helminth defence molecules (HDMs), which are similar to human antimicrobial peptides and might have both direct bactericidal and immunomodulatory effects^[62].

In this context, *G. lamblia* was found to interact directly with the intestinal microbiota, altering the immunopathology and infection outcomes^[63]. It is able to cause microbial dysbiosis with shifting towards virulent microbiota. The same was reported with *Blastocystis* spp. infection that was associated with a huge bacterial population in malnourished adults^[40]. On the other hand, it was suggested that the different microbiota inhibit parasitic infections by stimulating immune response through bacterial toll-like receptors^[6].

Attempts to comprehend the role of the human gut microbiome in childhood malnutrition, did not employ intestinal biopsy to characterize the microbiome^[64]. The mucosa-associated microbiome is widely thought to be more informative than fecal microbiota analysis, and future research will undoubtedly address this particular question^[64]. Three methods are suggested for manipulating microbiota: the use of antibiotics, probiotics (and related pre- and pro-synbiotics), and fecal transplantation^[56]. Apart from the role of gut commensal microbiota in the mechanisms that influence the survival and outcome of many parasitic infections, probiotics might be beneficial in lowering the pathogenicity of many parasites^[65]. Probiotics, according to WHO, are "live organisms that, when administered in adequate amounts, grant a health benefit to the host"^[66]. Probiotics have been shown to be effective in treating respiratory infections, allergic symptoms, and gastrointestinal disorders. They could also kill or inhibit pathogens via strain-specific mechanisms that involve competition, molecule

secretion, and/or immune induction^[66]. Accordingly, probiotics may be a factor in preventing the spread of several parasites, including some helminths like *A. lumbricoides* and *Trichuris* spp., and protozoans such as *Cryptosporidium* spp., and *G. lamblia*^[65].

Several lactobacilli species were found to have cytostatic and/or cytotoxic effects on *Giardia* trophozoites, both *in vitro* and *in vivo*. Other probiotic strains, demonstrated potent anti-*Giardia* properties both *ex-vivo* and *in vivo*, including *Weissella paramesenteroides*, *Bifidobacterium* spp., *Saccharomyces boulardii*, *Enterococcus faecium*, multi-strain probiotic, mixed bacteria, and yeast cultures as Slab51 and kefir grains^[67]. Furthermore, dietary prebiotics could be used to treat parasitic infections by promoting the growth of specific gut microbes that constrain or mitigate parasite virulence. Notably, simple dietary modifications might also be an effective anti-parasitic intervention^[6]. Hence it was suggested that new and innovative areas of research, such as the parasitome and metabolome of gut microbiota during chronic parasite infection, in addition to their relationship with host immunoregulatory mechanisms, should now be confronted in an integrated approach^[66].

CONCLUDING REMARKS

1. Malnutrition constructs a major public health issue in low resource settings. Underlying determinants are insufficient food intake, poor food hygiene, and recurring infectious diseases. Recurrent infections combined with poor nutrition creates a vicious cycle of increasing susceptibility to infection and worsening nutritional status, which in turn impacts child development in such a devastating way that might be irreversible.
2. Enteric parasitosis, in particular, is frequent in resource constrained regions known for high prevalence of different forms of malnutrition particularly, stunting. The means by which parasites can impact nutritional status include impairing of appetite, hindering intestinal absorption, increasing catabolism, and conveying nutrients away from growth. Some nutrient deficiencies also can hamper the host's immune response potentially lowering resistance to infections and rendering the host more vulnerable to infections. Additionally are the associated disruptions in intestinal absorptive and/or barrier function, with or without overt diarrhoea.
3. The EED is becoming a rising concern because of its impact on long-term public health issues. However, flaws in our understanding of its pathophysiology and its relationship to stunting limit ability to diagnose, prevent, and treat this condition efficiently. Furthermore, poor WASH conditions are increasingly regarded as a major contributor and are increasingly becoming the focus of targeted interventions aimed at improving nutritional status of children. Owing to the importance of intestinal microbiome in the development and maintenance of gut homeostasis,

its alteration might also contribute to both the onset and advancement of intestinal diseases. Accordingly, intestinal parasite carriage, micronutrient deficiency, EED, and alteration of gut microbiota interact through various pathways, that should be addressed concurrently to fight malnutrition among vulnerable pediatrics population in poor resource settings (Figure 1).

4. Hence, paving the way for implementing behavioural and therapeutic approaches are important factors for curing children with enteric parasitic infection and improving their nutritional status.
5. Suggested strategies should include routine screening for parasitic infections in the stool, anemia, and growth follow-up among children in primary health care units and schools; and administration of deworming medications. Vitamin A supplementation on regular basis would be valuable for both asymptomatic carriers and those with hidden nutritional issues. Water (advancements in both quantity and quality), sanitation (proper disposal of feces in particular),

and hygiene (education/promotion) interventions are also regarded as both helpful and beneficial tools in reducing undernutrition. Prescription of appropriate antibiotic therapy could overcome the chronic exposure to environmental microbes and pathogens that induce enteropathy. Furthermore, if malnourishment caused by helminthic infection triggers dysbiosis, supplements in the form of probiotics (and related pre- and pro-synbiotics) might be a beneficial therapeutic tool for the complementation of standard helminth treatment) (Figure 2).

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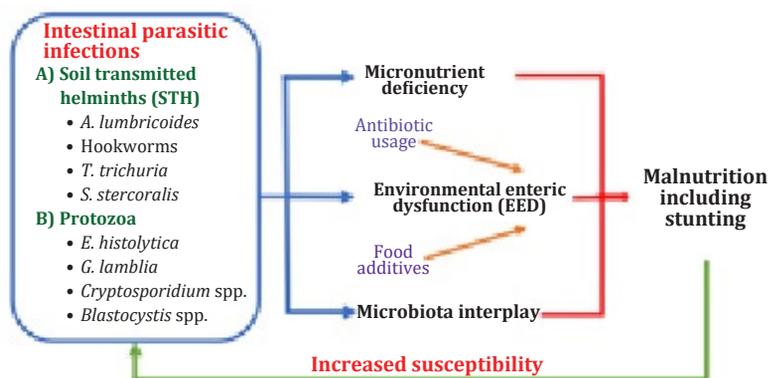


Fig. 1. Cumulative effect of repeated intestinal parasitic infections on children's nutritional status from different aspects. Illustrated by Zahran F.

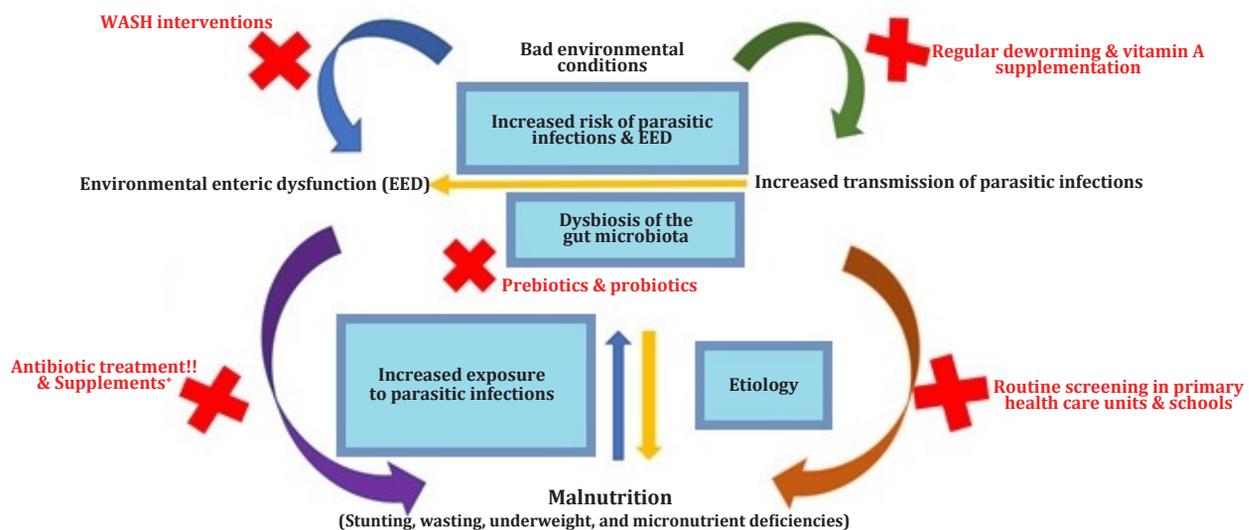


Fig. 2. Suggested interventions to overcome the cumulative effect of repeated intestinal parasitic infections on children's nutritional status. **WASH:** Water, sanitation, and hygiene; *****: How to interrupt the cycle?; *****: Probiotics and related pre- and pro-synbiotics. Illustrated by Zahran F.

REFERENCES

1. Kruk ME, Gage AD, Arsenuault C, Jordan K, Leslie HH, Roder-DeWan S, *et al.* High-quality health systems in the sustainable development goals era: Time for a revolution. *Lancet Glob Health* 2018; 6(11):e1196-e1252.
2. Akombi BJ, Agho KE, Merom D, Renzaho AM, Hall JJ. Child malnutrition in sub-Saharan Africa: A meta-analysis of demographic and health surveys (2006-2016). *PLoS One* 2017; 12(5):e0177338.
3. World Health Organization. Malnutrition. Available from <https://www.who.int/news-room/questions-and-answers/item/malnutrition>), Last update April 15, 2020.
4. Crane RJ, Jones KD, Berkley JA. Environmental enteric dysfunction: An overview. *Food Nutr Bull* 2015; 36(1): S76-87.
5. Prendergast AJ, Kelly P. Interactions between intestinal pathogens, enteropathy and malnutrition in developing countries. *Curr Opin Infect Dis* 2016; 29(3):229-236.
6. Leung JM, Graham AL, Knowles SC. Parasite-microbiota interactions with the vertebrate gut: Synthesis through an ecological lens. *Front Microbiol* 2018; 9:843.
7. Abbaszadeh Afshar MJ, Barkhori Mehni M, Rezaeian M, Mohebbali M, Baigi V, Amiri S, *et al.* Prevalence and associated risk factors of human intestinal parasitic infections: A population-based study in the southeast of Kerman province, southeastern Iran. *BMC Infect Dis* 2020; 20 (1):12.
8. Jourdan PM, Lamberton PH, Fenwick A, Addiss DG. Soil-transmitted helminth infections. *Lancet* 2018; 391(10117):252-265.
9. Hodges MH, Dada N, Warmsley A, Paye J, Bangura MM, Nyorkor E, *et al.* Mass drug administration significantly reduces infection of *Schistosoma mansoni* and hookworm in school children in the national control program in Sierra Leone. *BMC Infect Dis* 2012; 12:16.
10. Mekonnen HS, Ekubagewargies DT. Prevalence and factors associated with intestinal parasites among under-five children attending Woreta Health Center, Northwest Ethiopia. *BMC Infect Dis* 2019; 19(1):1-8.
11. Amare B, Ali J, Moges B, Yismaw G, Belyhun Y, Gebretsadik S *et al.* Nutritional status, intestinal parasite infection and allergy among school children in Northwest Ethiopia. *BMC Pediatr* 2013;13(1):1-9.
12. Korpe PS, Haque R, Gilchrist C, Valencia C, Niu F, Lu M, *et al.* Natural history of cryptosporidiosis in a longitudinal study of slum-dwelling Bangladeshi children: Association with severe malnutrition. *PLoS Negl Trop Dis* 2016; 10(5):e0004564.
13. Bourke CD, Jones KDJ, Prendergast AJ. Current understanding of innate immune cell dysfunction in childhood undernutrition. *Front Immunol* 2019; 10:1728.
14. Rajoo Y, Ambu S, Lim YA, Rajoo K, Tey SC, Lu CW, *et al.* Neglected intestinal parasites, malnutrition and associated key factors: A population based cross-sectional study among indigenous communities in Sarawak, Malaysia. *PLoS One* 2017; 12(1):e0170174.
15. El-Sherbini GT, Abosdera MM. Risk factors associated with intestinal parasitic infections among children. *J Egypt Soc Parasitol* 2013; 43(1):287-294.
16. Pullan RL, Smith JL, Jasrasaria R, Brooker SJ. Global numbers of infection and disease burden of soil transmitted helminth infections in 2010. *Parasit Vectors* 2014; 7(1):1-9.
17. Galgamuwa LS, Iddawela D, Dharmaratne SD. Prevalence and intensity of *Ascaris lumbricoides* infections in relation to undernutrition among children in a tea plantation community, Sri Lanka: A cross-sectional study. *BMC Pediatr* 2018; 18(1):1-9.
18. Liu C, Luo R, Yi H, Zhang L, Li S, Bai Y, *et al.* Soil-transmitted helminths in southwestern china: a cross-sectional study of links to cognitive ability, nutrition, and school performance among children. *PLoS Negl Trop Dis* 2015; 9(6):e0003877.
19. Richard SA, Black RE, Gilman RH, Guerrant RL, Kang G, Lanata CF, *et al.* Diarrhea in early childhood: Short-term association with weight and long-term association with length. *Am J Epidemiol* 2013; 178(7):1129-1138.
20. Buzigi E. Prevalence of intestinal parasites, and its association with severe acute malnutrition related diarrhea. *JBAH* 2015; 5(2): 2224-3208.
21. Bartelt LA, Lima AA, Kosek M, Peñataro Yori P, Lee G, Guerrant RL. Barriers to child development and human potential: the case for including the neglected enteric protozoa (NEP) and other enteropathy-associated pathogens in the NTDs. *PLoS Negl Trop Dis* 2013; 7(4):e2125.
22. Gombart AF, Pierre A, Maggini S. A review of micronutrients and the immunosystem- working in harmony to reduce the risk of infection. *Nutrients* 2020; 12(1):236.
23. Weyh C, Krüger K, Peeling P, Castell L. The role of minerals in the optimal functioning of the immune system. *Nutrients* 2022; 14(3):644.
24. Mahmoud AM, Abdul Fattah M, Zaher TI, Abdel-Rahman SA, Mosaad N. Intestinal parasitic infections and iron deficiency anaemia among school children in El Khalige Village, Dakhalia, Egypt. *AEJI* 2017; 7(1):28-36.
25. Papier K, Williams GM, Luceres-Catubig R, Ahmed F, Olveda RM, McManus DP, *et al.* Childhood malnutrition and parasitic helminth interactions. *Clin Infect Dis* 2014; 59(2):234-243.
26. Bundy DA, Walson JL, Watkins KL. Worms, wisdom, and wealth: Why deworming can make economic sense. *Trends Parasitol* 2013; 29(3):142-148.
27. Hodge C, Taylor C. Vitamin A deficiency. In: *Stat Pearls*. Treasure Island (FL): StatPearls Publishing; May 15, 2022.
28. Clohossey PC, Katcher HI, Mogonchi GO, Nyagoha N, Isidro MC, Kikechi E, *et al.* Coverage of vitamin A supplementation and deworming during Malezi Bora in Kenya. *J Epidemiol Glob Health* 2014;4(3):169-176.
29. de Gier B, Campos Ponce M, van de Bor M, Doak CM, Polman K. Helminth infections and micronutrients in school-age children: A systematic review and meta-analysis. *Am J Clin Nutr* 2014; 99(6):1499-1509.

30. Al-Mekhlafi HM, Anuar TS, Al-Zabedi EM, Al-Maktari MT, Mahdy MAK, Ahmed A, *et al.* Does vitamin A supplementation protect school children from acquiring soil-transmitted helminthiasis? A randomized controlled trial. *Parasit Vectors* 2014; 7, 367.
31. Taylor-Robinson DC, Maayan N, Soares-Weiser K, Donegan S, Garner P. Deworming drugs for soil-transmitted intestinal worms in children: Effects on nutritional indicators, haemoglobin and school performance. *Cochrane Database Syst Rev* 2012(11), DOI: 10.1002/14651858.CD000371.pub5.
32. Schmidt CW. Beyond malnutrition: The role of sanitation in stunted growth. *Environ Health Perspect* 2014; 122(11):A298-303.
33. Korpe PS, Petri Jr WA. Environmental enteropathy: Critical implications of a poorly understood condition. *Trends Mol Med* 2012; 18(6):328-336.
34. Pinkerton R, Oriá RB, Lima AA, Rogawski ET, Oriá MO, Patrick PD, *et al.* Early childhood diarrhea predicts cognitive delays in later childhood independently of malnutrition. *Am J Trop Med Hyg.* 2016;95(5):1004-1010.
35. Platts-Mills JA, Babji S, Bodhidatta L, Gratz J, Haque R, Havt A, *et al.* Pathogen-specific burdens of community diarrhoea in developing countries: A multisite birth cohort study (MAL-ED). *Lancet Glob Health* 2015; 3(9):e564-575.
36. Garzón M, Pereira-da-Silva L, Seixas J, Papoila AL, Alves M, Ferreira F *et al.* Association of enteric parasitic infections with intestinal inflammation and permeability in asymptomatic infants of Sao Tome Island. *Pathog Glob Health* 2017; 111(3):116-127.
37. Chen TL, Chen S, Wu HW, Lee TC, Lu YZ, Wu LL *et al.* Persistent gut barrier damage and commensal bacterial influx following eradication of *Giardia* infection in mice. *Gut Pathog* 2013; 5(1):1-2.
38. King IL, Li Y. Host-parasite interactions promote disease tolerance to intestinal helminth infection. *Front Immunol* 2018; 9:2128.
39. Badparva E, Kheirandish F. *Blastocystis* hominis: A pathogenic parasite. *Arch Clin Infect Dis* 2020; 15(4):e97388.
40. Fahim SM, Gazi MA, Hasan MM, Alam MA, Das S, Mahfuz M, *et al.* Infection with *Blastocystis* spp. and its association with enteric infections and environmental enteric dysfunction among slum-dwelling malnourished adults in Bangladesh. *PLoS Negl Trop Dis* 2021; 15(8):e0009684.
41. Watanabe K, Petri Jr WA. Environmental enteropathy: Elusive but significant subclinical abnormalities in developing countries. *EBioMedicine* 2016; 10:25-32.
42. Uddin MI, Hossain M, Islam S, Akter A, Nishat NS, Nila TA *et al.* An assessment of potential biomarkers of environment enteropathy and its association with age and microbial infections among children in Bangladesh. *PloS One* 2021; 16(4):e0250446.
43. Amaruddin AI, Koopman JPR, Muhammad M, Lenaerts K, van Eijk HHM, Brienen EAT, *et al.* Intestinal permeability before and after albendazole treatment in low and high socioeconomic status schoolchildren in Makassar, Indonesia. *Sci Rep* 2022;12(1):3394.
44. Waller A, Lakhapaul M, Godfrey S, Parikh P. Multiple and complex links between babyWASH and stunting: an evidence synthesis. *J Water Sanit Hyg Dev* 2020; 10(4):786-805.
45. Headey D, Palloni G. Water, sanitation, and child health: Evidence from subnational panel data in 59 countries. *Demography* 2019;56(2): 729-752.
46. Humphrey JH, Mbuya MNN, Ntozini R, Moulton LH, Stoltzfus RJ, Tavengwa NV, *et al.* Independent and combined effects of improved water, sanitation, and hygiene, and improved complementary feeding, on child stunting and anaemia in rural Zimbabwe: A cluster-randomised trial. *Lancet Glob Health* 2019;7(1):e132-e147.
47. Gough EK, Moodie EE, Prendergast AJ, Johnson SMA, Humphrey JH, Stoltzfus RJ, *et al.* The impact of antibiotics on growth in children in low and middle income countries: Systematic review and meta-analysis of randomised controlled trials. *BMJ* 2014; 348:g2267.
48. Ramirez J, Guarner F, Bustos Fernandez L, Maruy A, Sdepanian VL, Cohen H. Antibiotics as major disruptors of gut microbiota. *Front Cell Infect Microbiol* 2020; 10: 572912.
49. Becattini S, Taur Y, Pamer EG. Antibiotic-induced changes in the intestinal microbiota and disease. *Trends Mol Med* 2016;22(6):458-478.
50. Marion-Letellier R, Amamou A, Savoye G, Ghosh S. Inflammatory bowel diseases and food additives: to add fuel on the flames!. *Nutrients* 2019; 11(5):1111.
51. Lerner A, Matthias T. Changes in intestinal tight junction permeability associated with industrial food additives explain the rising incidence of autoimmune disease. *Autoimmun Rev* 2015;14(6):479-489.
52. Yan H, Ajuwon KM. Butyrate modifies intestinal barrier function in IPEC-J2 cells through a selective upregulation of tight junction proteins and activation of the Akt signaling pathway. *PLoS One* 2017;12(6):e0179586.
53. Raoul P, Cintoni M, Palombaro M, Basso L, Rinninella E, Gasbarrini A, *et al.* Food additives, a key environmental factor in the development of IBD through gut dysbiosis. *Microorganisms* 2022; 10(1):167.
54. Iacob S, Iacob DG, Luminos LM. Intestinal microbiota as a host defense mechanism to infectious threats. *Front Microbiol* 2019; 9:3328.
55. Evans JM, Morris LS, Marchesi JR. The gut microbiome: the role of a virtual organ in the endocrinology of the host. *J Endocrinol* 2013; 218(3):R37-47.
56. Kane AV, Dinh DM, Ward HD. Childhood malnutrition and the intestinal microbiome. *Pediatr Res* 2015; 77(1):256-262.
57. Subramanian S, Huq S, Yatsunenkov T, Haque R, Mahfuz M, Alam MA, *et al.* Persistent gut microbiota immaturity in malnourished Bangladeshi children. *Nature* 2014; 510(7505):417-421.
58. Blanton LV, Charbonneau MR, Salih T, Barratt MJ, Venkatesh S, Ilkaveya O, *et al.* Gut bacteria that prevent

- growth impairments transmitted by microbiota from malnourished children. *Science* 2016; 351(6275):10.
59. Kristensen KH, Wiese M, Rytter MJ, Özçam M, Hansen LH, Namusoke H, *et al.* Gut microbiota in children hospitalized with oedematous and non-oedematous severe acute malnutrition in Uganda. *PLoS Negl Trop Dis* 2016; 10(1):e0004369.
60. Glendinning L, Nausch N, Free A, Taylor DW, Mutapi F. The microbiota and helminths: Sharing the same niche in the human host. *Parasitology* 2014; 141(10):1255-1271.
61. Osborne LC, Monticelli LA, Nice TJ, Sutherland TE, Siracusa MC, Hepworth MR, *et al.* Virus-helminth coinfection reveals a microbiota-independent mechanism of immunomodulation. *Science* 2014; 345(6196): 578-582.
62. Cotton S, Donnelly S, Robinson MW, Dalton JP, Thivierge K. Defense peptides secreted by helminth pathogens: Antimicrobial and/or immunomodulator molecules? *Front Immunol* 2012; 3:269.
63. Fink MY, Singer SM. The intersection of immune responses, microbiota, and pathogenesis in *Giardiasis*. *Trends Parasitol* 2017; 33(11):901-913.
64. Hodges P, Tembo M, Kelly P. Intestinal biopsies for the evaluation of environmental enteropathy and environmental enteric dysfunction. *J Infect Dis* 2021; 224(Suppl. 7):S856-863.
65. Berrilli F, Di Cave D, Cavallero S, D'Amelio S. Interactions between parasites and microbial communities in the human gut. *Front Cell Infect Microbiol* 2012; 2:141.
66. Travers MA, Florent I, Kohl L, Grellier P. Probiotics for the control of parasites: An overview. *J Parasitol Res* 2011; 2011:610769.
67. Fekete E, Allain T, Siddiq A, Sosnowski O, Buret AG. *Giardia* spp. and the gut microbiota: Dangerous liaisons. *Front Microbiol* 2021;11:618106.